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# OPHIOCELAENO SELLNICKI, A NEW GENUS <br> AND SPECIES OF DIPLOGYNIIDAE ASSOCIATED WITH SNAKES (Acari-Mesostigmata) ( ${ }^{1}$ ) 

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The mesostigmatid mites of the superfamily Celaenopsoidea (currently comprised of the families Celaenopsidae, Euzerconidae, Diplogyniidae, and Schizogyniidae) are typically associated with arthropods or free-living, as adults. A prominent exception to this ecological generalization is the schizogyniid, Indogynium lindbergi Sellnick, 1950, which was described from specimens collected from three genera of snakes in the Palni Mts. of southern India. The association of Indogynium with a reptile has been questioned by Strandtmann and Wharton (1958) who correctly point out that such an instance runs counter to what we know of the Celaenopsoidea. It must be admitted, however, that not much is neally known about celaenopsoid mites and, further, there is a parallel (and in this instance, undoubted) case in the Antennophoroidea in which a single genus, Ophiomegistus, is a parasite of snakes and

[^0]lizards although all other genera are associates of arthropods. This question of true vertebrate- association is again opened for the Celaenopsoidea by the appearance of a second lot of specimens associated with snakes. This material consists of II4 adult males and females, representing a new genus and species of Diplogyniidae, taken from Typhlops sp. in Bougainville. The specimens wene among a large lot of Acari received recently from Dr. H.W. Levi of the Museum of Comparative Zoology, Harvard University, and we are most grateful to Dr. Levi for this interesting material.

## DIPLOGYNIIDAE

## Ophiocelaeno n.gen. (4)

Idiosoma broadest at level of region between coxae III and IV; narrowed in region of opisthosoma; not acute posteriorly. Dorsal shield markedly heterotrichous; with all interior setae reduced. Marginal dorsal setae short, spine-like laterally (ro pairs) ; elongate posteriorly ( 5 pairs). IWith 3 pairs of elongate submarginals. No dorsal neotrichy. Sternal shield of female relatively long and with sternal setae II close together ; sternals I and III well separated ; all sternal setae at separate levels. Latigynial shields not excavated anteriorly; with 2 pairs of setae. Ventral and anal shields fused; no ventral neotrichy. Chelicerae with digits relatively narrow ; with many small, closely-set teeth. Male with laterally flattened, spoon-shaped process on fixed digit and horn-like process on disti-ventral margin of cheliceral base. Tectum with median rib reduced in anterior half. All legs relatively short, stout.

Type species: Ophiocelaeno sellnicki n.sp.

## Ophiocelaeno sellnicki n.sp. (5)

Female. Idiosoma rounded anteriorly, with moderate shoulders; widest at level between coxae III and IV and narrowed posteriorly. Length of idiosoma $789-853 \mu$ in II specimens measured.

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Fig. x. - Dorsal view of idiosoma of female.


Fig. 2. - Ventral view of idosoma of female.

Dorsum. (Figure I) Dorsal shield markedly heterotrichous and with complex porotaxy. Forty-two pairs of dorsal setae present in addition to 1 unpaired (and often assymetrically placed) seta posterior to $i_{1}$. Setal complement consisting of 24 pairs of short


Fig. 3. - Sterni-genital region of female.
or vestigial setae; 3 pairs of long, curved submarginal setae; 15 pairs of marginal setae of which posterior 5 pairs are enlarged.

Venter. (Figures 2 and 3) Tritosternum normal; with 2 spinose laciniae. Sternal shield with thickened anterior margin. Sternal
setae I long, spine-like; reaching to level of sternal setae II. Sternals II small; in center of sternal shield; about one-third as far apart as sternals I. Sternals III short; about the same distance apart as sternals I. Metasternal shields free; setae very small. Latigynial shields with antero-lateral margins concave, not incised; with 2 pairs of setae. Latigynials I approximately in center of shield; latigynials 2 on median margin just anterior to the small, triangular mesogynial shield. Ventral shield fused with peritremal-parapodal complex and with anal shield; V-shaped, with slightly convex sides; reaching almost to posterior margin of opisthosoma. Ventral shield bearing 4 pairs of ventral and 2 pairs of paranal setae. Ventri-lateral shields each with long, sigmoid ridge in the posterior half; a pair of setae at the hind margin. Anus surrounded by a group of branched integumentary (? glandular) canals.

Gnathosoma (6). (Figures 4 and 5) Tectum triangular in outline ; sides smooth and gently concave ; extending almost to distal end of palp femur ; median rib restricted to basal half. Subcapitulum large ; well sclerotized. Hypotome well developed, reaching past level of palp trochanter; distally expanded and bifurcate. " Hypopharyngeal " processes filamentous, smooth, centrally ribbed; long, reaching past level of palp genu. Hypostomal processes present, originating just mediad of corniculi; reaching almost to level of tip of corniculus. Corniculi heavy, saber-like; reaching to or slightly beyond level of distal margin of palp trochanter. Hypostomal setae in oblique row ; lightly serrate; decreasing slightly in length from anterior to posterior pair. Subcapitular setae serrate; subequal in length with posterior hypostomals. "Deuterosternal w column with (modal) 3 finetooth files. Labrum (epipharynx of Gorirossi) finely spinose dorsally; smooth ventrally. Cheliceral digits (Figure 5) relatively long, thin; with many small, closely-set teeth in addition to basal locking teeth. Pilis dentilis vestigal. Dorsal seta of chelicera not observed. Movable digit with 2 tree-like excresences and a single saber-like excresence with small setules. Chaetotaxy of palp trochanter, femur, genu and tibia: 2-5-7-15. Pretarsus with 2 claws.

Legs relatively short; II-IV quite stout. Pretarsus present on

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Fig. 4. - Ventral view of gnathosoma of female.
legs II-IV; lacking on leg I. Chaetotaxy of legs as follows (using the system of Evans, 1963):

|  | I | II | 111 | IV |
| :---: | :---: | :---: | :---: | :---: |
| coxa | 2 | 2 | 2 | 1 |
| troch | I-I/3-1 | 5 | 5 | 5 |
| FEMUR | 1-2/I; $2 / 3$-1 | 2-2/土; 2/2-I* | 1-2/1; $2 / \mathrm{I}-\mathrm{O}$ | 1-2/r; $2 / \mathrm{I}-\mathrm{I}$ |
| genu | 1-3/1; 2/I-1 | I-3/I; 2/I-I | I-2/I; $2 / \mathrm{I}-\mathrm{I}$ | 1-2/I; 3/I-I** |
| TIBIA | 2-3/2; 2/2-2 | I-I/I; $2 / \mathrm{I}-\mathrm{I}$ | I-I/r; $2 / \mathrm{T}-\mathrm{I}$ | I-I/2; $2 / \mathrm{I}-\mathrm{I}$ *** |
| TARSUS**** | - | 19 | 19 | 20 |

* $A L_{2}$ and $\mathrm{PV}_{2}$ on basifemur.
** $\mathrm{PD}_{1}$ is a mastiseta.
* PV is stout, heart-shaped.

Count includes a prorals».
Females contained a maximum of 4 eggs.
Measurements of holotype female. Length of idiosoma $821 \mu$. Maximum width of idiosoma $581 \mu$. Distance between centers of bases of sternal setae I $88 \mu$; sternals II $28 \mu$; sternals III $93 \mu$. Length of sternal shield along median line $73 \mu$. Maximum length of latigynial shield $118 \mu$; maximum width $83 \mu$. Length of tarsus I I3I $\mu$.
Male. Length of idiosoma $736-853 \mu$ in 8 specimens measured. Dorsum as in female.
Venter (Figure 8) normal. Genital aperture at anterior margin of sternal shield. «Glandular " porose area in center of sternal shield behind genital aperture. Peritremal, parapodal, sternal, ventral, and anal shields fused; separated by ablique suture from ventri-lateral shields. Six pairs of sterni-genital setae present (ST I-IV ; G I-2) and 4 pairs of ventrals (V I-4) and 2 pairs of paranals ( $\mathrm{PA} \mathrm{I}_{\mathrm{I}-2}$ ) ; total complement as in female.

Gnathosoma similar to that of female except in following features: Corniculi (Figure 7) long, curved, setiform; reaching past tip of hypostome. Second hypostomal process present; arising antero-mediad of more lateral process. Chelicerae (Figure 6) with but a single, distally bifurcate treelike excresence and with a blunt, laterally compressed, spoon-shaped process on fixed digit. Distal portion of cheliceral base with ventral, horn-like, hyaline process.

Legs similar to those of female except that trochanter IV bears a prominent, semi-circular, scale-like process (present but less developed on trochanters II and III).


Fig. 5-6. - Chelicerae of female (5) and male (6).

Holotype female deposited in the U.S. National Museum, Washington, and bearing the following data: Ex Typhlops sp. (spotted type only) : Solomon Islands: Bougainville Isl. : Kieta: Nov. 1960: Fred Parker. Paratype males and females (all with same data as holotype) deposited in the British Museum (Nat. Hist.), London; the U.S. National Museum ; the Museum of Comparative Zoology, Cambridge ; the Snow Museum, University of Kansas, Lawrence ; the Musee d'Histoire Naturelle, Paris ; the Institut de Medecine Tropicale, Anvers; and the Institute of Acarology, Wooster.


Fis. 7. -- Partial ventral view of gnathosoma of male showing corniculus.

## DISCUSSION

In addition to those cited in Baker and Wharton, 1952 (whose list is based on the partial monograph of Trägàrdh, 1950), the following genera have been referred to the Diplogyniidae: Neolobogynium Hicks, 1957 (type species: N. americana (Banks, 1906) $=$ lateriseta Hicks, 1957 ; see Johnston 1960) ; Lobogyniella Krantz, 1958 (type species: L. tragardhi Krantz, 1958) ; Monodiplogynium Womersley, 1958 (type species: M. carabi Womersley, 1958) ; Paradiplogynium Womersley, 1958 (type species: $P$. panesthia Womersley, 1958) ; and Crenamargo Hicks, 1958 (type species: C. binuseta Hicks, 1958). There are also recent short papers on Diplogyniidae by Samsinak, 1957; Fox, 1957; and Hicks, 1959.


Fig. 8. -- Ventral view of idiosoma of male.

In this paper we shall make no attempt to compare Ophiocelaeno with other named genera of Diplogyniidae other than through the presentation in the generic diagnosis of that combination of features by which the new genus differs from all others. The family Diplogyniidae, as with other celaenopsoid groups, seems not so amenable to routine taxonomic procedures, and analysis of the group by the techniques of numerical taxonomy appears a requisite in dealing with the classification of these mites. Such an analysis is being undertaken by R. Funk and J.H. Camin of the University of Kansas.

Our study of Ophiocelaeno sellnicki has not revealed any features which can be interpreted with certainty as correlated with parasitism or any other type of association with the snake host. The cheliceral digits are relatively longer and narrower than in many diplogyniids and the hypostome is more elaborately developed than usual, but an interpretation of the significance of these features must await a comparative study of the family.

> ADDITION TO FAIN'S (I962) LIST OF MESOSTIGMATID MITES ASSOCIATED WITH SNAKES

## SCHIZOGYNIIDAE

Indogynium lindbergi Sellnick, 1954.
Ex Platyplectrurus madurensis Bed., Uropeltis pulnejensis (Bed.), and Teretrurus rhodogaster (Wall.) in Kodikanal, Palni Mts., India (Sellnick, 1954).

## REFERENCES CITED

Baker. E.W. and Wharton, G.W., 1952, An Introduction to Acarology. (New-York. i-xiii + I-465).
Evans, G.O., 1963, Observations on the chactotaxy of the legs in the freeliving Gamasina (Acari: Mesostigmata). (Bull. Brit. Mus. (Nat. Hist.), Zool., 10 (5): 275-303).
Fain, A., ig62, Les acariens mesostigmatiques rectoparasites des serpents. (Bull. Inst. Royal Sci. Nat. Belgique XXXVIII (18): I-149).
Fox, I., 1959, A new mite taken with rats in Puetto Rico (Acarina : Diplogyniidae) (Acarologia I (3): 296-298).
Gorirossi, F.E., I955, The gnathosoma of the Celaenopsina, )Acarina Mesostigmata) (Amer. Midl. Nat. 54 (1): 153-167).
Hicks, E.A., 1957, A new genus and species of the family Diplogyniidae (Acarina) (Proc. Iowa Acad. Sci. 64: 614-620).
Hicks, E.A., 1958, A new genus and species of diplogyniid from Nicaragua (order Acarina, family Diplogyniidae) (Iowa State Journ. Sci. 33 (2): 103-110).

Hicks, E.A., 1959, The male of Lobogynioides obtusum Trägårdh (Acarina, Diplogyniidae) (Proc. Iowa Acad. Sci. 66: 474-476).
Johnston, D.E., ig60, Some new synonymy in the Haemogamasidae, Laelaptidae and Diplogyniidae indicated by an examination of Banks' types of Mesostigmata (Acarina) (Psyche 66 (4): 60-62).
KRANTZ, G.W. I958, Lobogyniella tragardhi, a new genus and species of diplogyniid mite associated with dampwood termites in Oregon (Acarina, Diplogyniidae) (Proc. Ent. Soc. Wash. 60 (3): 127-13I).
Samsinak, K., I957, Die mitteleuropäischen Arten der Familie Diplogyniidae (Acari) (Acta Soc. Ent. Cechosloveniae 54 (I): I-6).
Sfllnick, M., I954, Indogynium lindbergi nov. gen., nov. spec., eine nele Acaride aus Indien (Ent. Tidskr. 75 (2-4): 285-291).
Strandtmann, R.W. and Wharton, G.W., I958, A Manual of Mesostigmatid Mites Parasitic on Vertebrates (Contrib. No. 4, Institute of Acarology. i-xi, 1-330, pls 1-69).
Trägardi, I., 1950, Studies on the Celaenopsidae, Diplogyniidae and Schizogyniidae (Acarina) (Ark, f. Zool., Ser. 2, I (25): 36I-45I).
Womersley, H., 1958, On some Acarina from. Australia and New Guinea paraphagic upon millipedes and cockroaches, and on beetles of the family Passalidae (Trans. Royal Soc. South Austral. 8r: 13-29).


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[^1]:    (4) Because of the lack of comparative studies in the Diplogyniidae, this and all previous generic diagnoses should not be taken too seriously.
    (5) This species is named for our friend and colleague, Dr. Max Sellnick (Deutschland).

[^2]:    (6) The terminology applied to the gnathosoma follows, in general, that of Gorirossi (1955).

